



OPEN ACCESS

*CORRESPONDENCE

Tammy R. Dugas,
✉ tammydugas@lsu.edu

RECEIVED 29 July 2025

REVISED 31 October 2025

ACCEPTED 18 November 2025

PUBLISHED 08 December 2025

CITATION

Akers NM and Dugas TR (2025)
Peripheral artery disease and local drug
delivery: a review of disease pathology
and drug delivery systems for therapy
below the knee.

Exp. Biol. Med. 250:10754.

doi: 10.3389/ebm.2025.10754

COPYRIGHT

© 2025 Akers and Dugas. This is an
open-access article distributed under
the terms of the [Creative Commons
Attribution License \(CC BY\)](#). The use,
distribution or reproduction in other
forums is permitted, provided the
original author(s) and the copyright
owner(s) are credited and that the
original publication in this journal is
cited, in accordance with accepted
academic practice. No use, distribution
or reproduction is permitted which does
not comply with these terms.

Peripheral artery disease and local drug delivery: a review of disease pathology and drug delivery systems for therapy below the knee

Nicole M. Akers and Tammy R. Dugas*

Department of Comparative Biomedical Sciences, Louisiana State University School of Veterinary
Medicine, Baton Rouge, LA, United States

Abstract

Peripheral artery disease (PAD) is a disease of both atherosclerotic and thromboembolic pathology, affecting more than 230 million people globally. PAD patients are at an increased risk of thrombotic events and often require lifelong antithrombotic therapy. Thromboembolism can lead to complete occlusion of affected arteries and put patients at risk for critical limb threatening ischemia (CTLI). PAD blockages are cleared using drug-eluting stents (DES) and drug-coated balloons (DCB). However, PAD treatment below the knee (BTK) presents unique challenges. While DCB are frequently used to treat BTK disease, no DCB has gained FDA approval for this indication. However, innovation in the field has produced drug delivery systems and formulations that may yet enhance the effectiveness of these therapies. In this review, we will provide a brief overview of the pathological mechanisms associated with PAD and review the materials and drugs frequently used in DCBs with an emphasis on excipients and drug carriers. Finally, we will highlight emerging devices undergoing clinical trials to treat BTK disease and how they differ from their predecessors.

KEYWORDS

peripheral artery disease, drug coated balloon, paclitaxel, sirolimus, thrombosis, combination device, biocompatibility

Impact statement

We provide timely updates to the progress being made in combination device development for peripheral artery disease (PAD) therapy. This review article summarizes both basic pathophysiologic information for PAD as well as device development considerations for combination devices. Lesions below the knee have proven challenging to treat. Drug coated balloons are frequently used as a part of PAD lesion treatment below the knee, yet none are approved for use below the knee in the US. Therefore, we discuss the latest updates in the development of several promising

combination and lesion preparation devices for treatment of PAD disease below the knee, a historically recalcitrant area to treat. This information will be useful to both scientists and clinicians who are either developing their own combination devices or looking for cutting edge information on how new devices are different from their predecessors.

Introduction

Peripheral artery disease (PAD) encompasses atherosclerotic and thrombotic pathology outside of the coronary and cerebral vascular systems. The most common presentation of PAD occurs within the lower limbs, with an estimated global prevalence of more than 230 million cases [1]. PAD is associated with significant morbidity, disability, and mortality in affected individuals. These subjects can experience limb weakness and claudication due to decreased tissue perfusion from narrowed, damaged vessels, up to complete occlusion of blood flow, leading to critical limb-threatening ischemia (CLTI) and potential limb amputation. The risk for myocardial infarction (MI) or stroke in PAD patients is on par with patients suffering from coronary artery disease [2]. PAD treatment includes a combination of lifestyle modifications, medical therapy, and when needed, endovascular interventions including surgical approaches and medical device interventions [3, 4]. There are a wide variety of devices available to treat PAD lesions, including the use of bare-metal stents (BMS) and drug-eluting stents (DES), either balloon-expanded or self-expanding, newer woven and covered nitinol stents, dissolvable scaffolds, percutaneous transluminal angioplasty (PTA) with either drug-coated (DCBs) or plain balloons (POBA), intravascular lithotripsy and atherectomy to treat calcified lesions [5]. Treatment approaches are highly dependent on the location, length and number of lesions present, as well as the pattern of disease in the individual and their comorbidities. While both stenting and balloon angioplasty have been successful above the knee, lesions below the knee (BTK) have restenosis rates that approach 70 percent [6]. Additionally, BTK lesions are heavily calcified, cover extensive lengths of the artery, and importantly, possess significant thromboembolic pathology that can lead to adverse outcomes including CLTI and subsequent amputation [7]. Below the knee, DCBs are commonly used to treat lesioned arteries; yet none are FDA approved for use below the knee due to a lack of evidence of long term benefits over POBA. Recent developments in DCB include novel drug formulations and carriers, which may yet improve clinical outcomes in the long term for BTK disease. These carriers include liposomal formulations, polymeric microspheres, and

aqueous delivery systems, among others. Before delving into these novel technologies, we will first discuss PAD pathophysiology, drugs commonly used in DCB, the difficulty associated with BTK disease treatment, and how coating formulations can enhance or derail effective DCB treatment.

Pathologic mechanisms of PAD

Endothelial regulation of thrombosis

Healthy endothelial cells express both prostacyclin (PGI_2) and endothelial nitric oxide synthase (eNOS). eNOS provides a source of nitric oxide (NO), which along with PGI_2 , synergistically inhibits platelet adhesion and aggregation via binding to receptors expressed on the platelet surface, reducing their activity [8]. NO is additionally a vasodilator that permeates the endothelium, promoting relaxation of the vascular smooth muscle. Vascular endothelial cells are key regulators of coagulation and thrombosis. TV-VIIa (activated factor VII complex) and prothrombinase are key initiators of early clot development; vascular endothelial cells express TFPI- α and TFPI- β (tissue factor pathway inhibitor alpha and beta, respectively) that inhibit the TF-VIIa (activated factor VII) complex and prothrombinase. Therefore, TFPI- α and TFPI- β inhibit clot formation at an early stage [9]. Fibrinolysis is regulated via plasminogen activator inhibitor (PAI-1), endothelial urokinase plasminogen activator (u-PA) and tissue plasminogen activator (t-PA). Thus, under normal circumstances the luminal endothelial surface is antithrombogenic, expressing multiple inhibitors of coagulation, platelet aggregation and adhesion, as well as other factors that promote fibrinolysis. On the other hand, decreased eNOS activity in damaged endothelia promotes platelet aggregation, reactivity, and thrombosis [8].

Invasion of lipids and inflammatory cells

Inflammatory cytokines, reactive oxygen species, and high levels of circulating LDLs behave as endothelial stressors [10, 11]. Under chronic exposure to stressors, endothelia can become dysfunctional. Dysregulated eNOS activity due to endothelial dysfunction reduces NO output. NO is critical for maintaining endothelial barrier function, and NO inhibits NF- κ B, a key transcription factor that promotes the expression of ICAM, VCAM-1, E-selectin, and other leukocyte adhesion receptors on the endothelium [12]. As a result of compromised endothelial barrier function, the endothelium becomes permeable to the transmigration of inflammatory cells and lipids via decreased NO production [13]. Monocytes thus

transmigrate through the endothelium and differentiate into macrophages, phagocytizing LDLs. Phagocytosis of LDL transforms macrophages into foam cells. These foam cells become apoptotic and are cleared by M2 macrophages. When these macrophages die, they release TF, lipids, and other inflammatory molecules including matrix metalloproteinases (MMP), further promoting a prothrombotic and inflammatory state [14].

Atheroma development

Transformation of macrophages into foam cells that deposit lipids into the subendothelial matrix marks the initiation of atheroma development. Like coronary artery disease (CAD), atherosclerotic plaques can be found in PAD lesions. Atherogenesis can be defined by several stages, beginning with a fatty streak within the artery wall, progressing to a fibrous plaque, to an unstable plaque at risk of rupture, and ultimately, a ruptured plaque. Atherosclerotic plaques contain a mixture of lipids, minerals, inflammatory cells, platelets and cellular degradation products [15]. Additionally, ruptured atherosclerotic plaques release pro-thrombotic factors such as tissue factor (TF) from the necrotic core, which promotes thrombosis [16].

Medial calcification

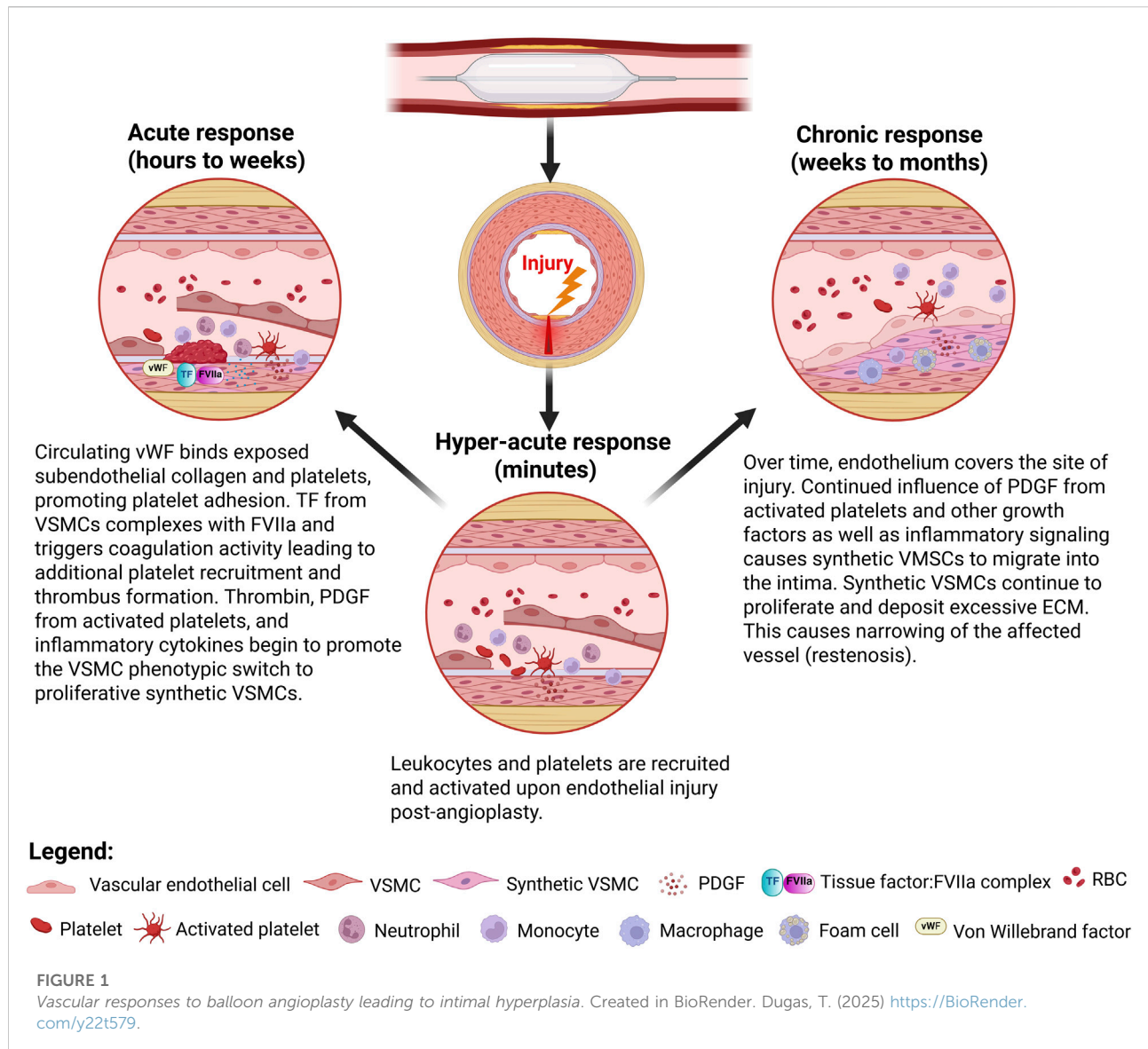
Calcification associated with CAD and calcified lesions associated with PAD differ. Calcification can be classified as intimal or medial depending on where it is located within the vessel tunics, each with different proposed mechanisms for their pathogenesis. While CAD calcification is primarily intimal and associated with fibrous plaques, PAD calcification can be found in both the intima and media [17]. Importantly, medial calcification is often observed with CTLI [7]. Medial calcification was once considered a benign, aging-associated change in the tunica media; however, it may contribute to thrombotic risk. While atherosclerotic plaque rupture is considered a central mechanism for thromboembolism in the presence of plaques, highly calcified BTK lesions with minimal atherosclerotic involvement also display thromboembolic pathology [7]. This suggests the possibility that there are alternative pro-thrombotic mechanisms outside of atherosclerotic plaque rupture. Chang and coworkers proposed lower leg arterial calcification as a potential risk factor for acute thrombosis independent of atherosclerotic pathology [18]. Accordingly, luminal thrombi in BTK lesions are not routinely associated with atherosclerotic plaques compared to CAD lesions but are commonly associated with heavily calcified arteries [19]. Narula et al., examined 299 arteries collected from

95 patients with CTLI. BTK arteries presented more often with diffuse, chronic, occlusive thromboembolic pathology and significant calcification with minimal atherosclerotic disease when compared to FP lesions, although both BTK and FP segments displayed medial calcification [7].

Additionally, different patterns of calcification present in FP and BTK lesions may complicate patient outcomes. FP segments often have thick, patchy calcifications associated with the tunica intima, whereas BTK lesions frequently present with continuous, annular calcifications which form a circumferential ring of arterial calcification [20]. Annular calcification patterns have been tied to poor long term survival in patients with CTLI [21]. Lastly, intensely calcified arteries pose a significant physical barrier against the transfer of drugs through the arterial wall.

Mechanisms of vascular remodeling following balloon injury in PAD patients

Balloon angioplasty has become the preferred approach to BTK lesions. However, endothelial damage including complete denudation of the endothelial layer at the site of angioplasty can occur [22]. This damage contributes to IH development, which starts with the recruitment of platelets and inflammatory cells to injured sites and results in the synthetic vascular smooth muscle cells (sVSMC) phenotypic switch that promotes remodeling of the arterial wall. Circulating platelets adhere to exposed subendothelial collagen by binding to vWF and collagen via platelet surface glycoproteins GPVI and GPIb-IX-V [23]. Bound platelets become activated and recruit additional platelets through the release of aggregatory mediators including ADP, thromboxane A₂, and thrombin [24]. Thrombin generated through TF-mediated extrinsic coagulation activity further bolsters platelet aggregation [25]. Importantly, these bound platelets secrete platelet-derived growth factor (PDGF), a potent activator of VSMC migration and proliferation [26, 27]. Additionally, macrophages have been long established as promoters of the sVSMC phenotype. M1 macrophages, which are often found in association with atherosclerotic lesions and in damaged tissues following balloon angioplasty, secrete numerous cytokines including TNF- α , IL-1 β and IL-6 [28]. These cytokines have all been implicated in modulating the VSMC phenotype, enhancing proliferation and migration [29, 30]. Under the influence of growth factors PDGF as well as inflammatory cytokines, VSMC behave as sVSMC. These sVSMC migrate from the tunica media to the tunica intima, where they proliferate and secrete extracellular matrix (ECM) components leading to ECM deposition and expansion [31]. Ultimately, the root of the remodeling process is endothelial injury triggering this cascade of events [Figure 1](#).



Challenges in treating PAD below the knee

Drug coated devices targeting vascular smooth muscle cell proliferation have improved vascular patency over bare metal stents and uncoated balloons in femoropopliteal lesions [32, 33]. However, long term benefits from DCBs in BTK segments have yet to be established. Below the knee, the infrapopliteal vessels including the anterior and posterior tibial, fibular, and pedal arteries are smaller, thinner, and are exposed to numerous mechanical forces. Stents run an elevated risk of fracture BTK and are typically used as a bailout option. Balloon angioplasty is considered the primary therapy to treat BTK PAD. However, there are multiple success-limiting factors for BTK lesions which include long lesion length, significant calcification, vascular

elastic recoil, flow-limiting dissection, and restenosis [34]. While many DCB have gained FDA approval, none are approved for BTK arteries Table 1.

Drugs used in endovascular device coatings and their cellular targets

Paclitaxel

Paclitaxel, a potent tumoricidal drug, was first isolated in 1967. Paclitaxel has been used extensively as a cancer therapy. However, research performed in the 1990s demonstrated the ability of paclitaxel to inhibit VSMC proliferation [35]. The first paclitaxel-eluting stent was approved by the FDA for use in the

TABLE 1 Endovascular combination devices approved by the FDA in the past 10 years for the treatment of PAD.

Device	Manufacturer	FDA approval status	Lesion indication	Drug (µg/mm ²)	Coating
Esprit™ BTK everolimus eluting resorbable scaffold system	Abbott vascular (IDEF technologies inc.)	Approved 4/26/24	BTK	Everolimus 1 µg/mm ²	Poly (D,L-lactide)
SurVeil drug-coated balloon	Surmodics, inc.	Approved 6/16/23	FP	Paclitaxel 2 µg/mm ²	Polyethyleneimine polymer
Chocolate touch paclitaxel drug-coated PTA balloon catheter	TriReme medical, LLC, (now genesis medtech)	Approved 11/04/22	FP	Paclitaxel 2.95 µg/mm ²	Propyl gallate
Ranger™ paclitaxel-coated PTA balloon catheter	Boston Scientific corporation	Approved 10/30/20	FP	Paclitaxel 2 µg/mm ²	Acetyl tributyl citrate
Eluvia drug-eluting vascular stent system	Boston Scientific corporation	Approved 9/18/18	FP	Paclitaxel 0.167 µg/mm ²	PBMA (poly (n-butylmethacrylate)) and PVDF-HFP (vinylidene fluoride and hexafluoropropylene copolymer)
Stellarex 0.035 OTW drug-coated angioplasty balloon	The spectranetics corp.	Approved 7/26/17	FP	Paclitaxel 2 µg/mm ²	PEG-8000
IN.PACT admiral paclitaxel-coated PTA balloon catheter and IN.PACT 018 paclitax	Medtronic inc.	Approved 05/29/14	FP	Paclitaxel 3.5 µg/mm ²	Urea
Lutonix drug coated balloon PTA catheter	Lutonix	Approved 10/09/14	FP	Paclitaxel 2 µg/mm ²	Polysorbate and sorbitol

coronary arteries in 2004 after promising results from the TAXUS trials [36]. However, concerns regarding permanent stents and their association with IH lead to the development of paclitaxel coated balloons for coronary angioplasty. In the 2010s, paclitaxel-coated balloons underwent clinical trials to evaluate their use in small coronary arteries [37]. In 2015, the first trial using a Paclitaxel-coated DCB in FP arteries followed [38]. Since then, paclitaxel has been used commonly as a DCB coating. Paclitaxel acts as a microtubule stabilizing agent that prevents the tubular migration necessary for mitotic spindle assembly and causes cell cycle arrest in the G2/M phase [39]. Ultimately, these arrested cells undergo apoptosis. Paclitaxel upregulates BCL-2, DAP3, BAX, DAD1, and several other pro-apoptotic genes [40]. Additionally, paclitaxel affects multiple pathways associated with cellular proliferation, including receptor tyrosine kinases (RTK), TGF-β, and upstream regulators of the ERK pathway [40]. The effects of paclitaxel are non-specific; while paclitaxel inhibits VSMC proliferation, it may delay reendothelialization of denuded epithelia [41].

Paclitaxel controversy

With respect to decreased IH and reduced restenosis rates, the use of paclitaxel-coated devices represents a significant improvement over BMS and POBA. However, the use of paclitaxel in DES and DCB has not been without controversy. Katsanos et al. conducted a meta-analysis including 28 research-controlled trials assessing the use of paclitaxel-eluting DES and

DCB. Their study demonstrated an increased risk of mortality associated with paclitaxel-coated devices [42]. A follow up meta-analysis demonstrated no significant increase in all-cause mortality between 1 and 2 years, but an increased risk of mortality between year 3 and 5 [43]. In 2019, the FDA issued a letter to healthcare providers to notify them of the increased late mortality signal. Following this, manufacturers of FDA-approved devices submitted deidentified individual patient data to the VIVA Physicians medical research organization, which produced an aggregate meta-analysis published in Circulation in May 2020 stating that no increased mortality signal was found [44]. Further meta-analyses of clinical trial data found no increase in all-cause mortality [45]. The FDA also analyzed data from several trials including the VOYAGER PAD study, the BARMER Health Insurance study, the Medicare Safe-PAD study, the U.S. Veterans Health Administration study, and the SWEDEPAD interim analysis [46]. By July 2023, the FDA issued updated guidance stating there was no increased mortality signal.

Sirolimus

Sirolimus, also known as rapamycin, is a macrolide antibiotic with poor antibacterial capabilities that is used as an immunomodulatory, cytostatic, and antiproliferative agent. Sirolimus acts to inhibit the mTOR pathway by reversibly binding to FK506-binding protein 12 (FKBP12). FKBP12 binds tacrolimus (FK506) as well as rapamycin and

TABLE 2 Paclitaxel versus sirolimus: Reported effects exerted on cellular events that follow endovascular interventions.

Cellular events that follow intervention with stents and balloons	Paclitaxel	Sirolimus
Platelet aggregation	Inhibits collagen-mediated platelet aggregation and TXA ₂ synthase [49]; enhances platelet aggregation via increased sensitivity to ADP [50]	Enhanced platelet aggregation via increased platelet sensitivity to ADP [51, 52]
Inflammation	Increased [53, 54]	Reduced [55, 56]
VSMC proliferation	Inhibited [35]	Inhibited [57]
VSMC migration	Inhibited [35]	Inhibited [58]
Cell death	Promotes apoptosis and autophagy [39, 40, 59]	Prolongs the G1 phase prior to the G1/S checkpoint in a reversible manner [47]
Reendothelialization	Inhibited [60]	Inhibited [61]

other rapalogs, creating a complex that inhibits MTORC1 [47]. The downstream result of mTORC1 inhibition is that cells cannot progress through the G1/S transition and are maintained in G1. Sirolimus inhibits VSMC proliferation via this mechanism. Additionally, inhibition of NF- κ B by sirolimus has been previously demonstrated [48]. Inhibition of NF- κ B has downstream effects on the expression of leukocyte adhesion molecules and chemoattractants, conferring an anti-inflammatory role in addition to its other effects. While sirolimus and paclitaxel are some of the most frequently used drugs in DCBs, there are key differences in how they affect cellular processes Table 2.

Everolimus and other limus drugs

Everolimus, 40-O-(2-hydroxyethyl)-rapamycin, is a sirolimus analog with a hydroxyethyl group at C-40 [62]. Modification of sirolimus in this manner was intended to improve oral bioavailability but resulted in several key differences in the behavior of everolimus [63]. Everolimus is an mTOR inhibitor with a weaker binding affinity for FKBP12 than sirolimus. Unlike sirolimus, which only inhibits MTORC2 with chronic use, everolimus demonstrates activity against both MTORC1 and MTORC2 [64]. Similar to sirolimus, the end result is that cells do not progress past G1 of the cell cycle [65].

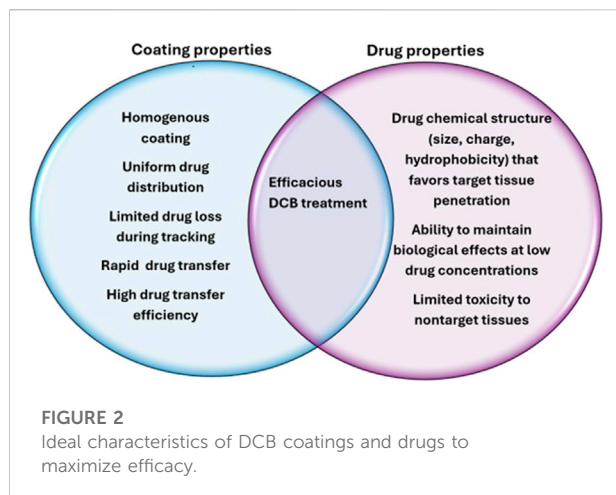
Everolimus was initially developed for use in solid organ transplants. Owing to its antiproliferative properties, interest in its use in coronary stents culminated in the FUTURE and SPIRIT FIRST clinical trials [66, 67]. In 2009, the FDA approved the first everolimus-eluting stent for use in the coronary arteries. Since then, everolimus-eluting stents have been designed for use in peripheral arteries, including the XIENCE Prime™ BTK Everolimus Eluting Peripheral Stent by Abbott (currently marketed outside of the US). Most recently, the Espirit™ BTK everolimus eluting resorbable scaffold system by Abbott Vascular was approved by the FDA for use in infrapopliteal arteries.

Zotarolimus is the first limus drug designed specifically for use in drug-eluting stents. Some of these stents include the Medtronic Endeavor stents and now the Medtronic Onyx Frontier stent, which are used in CAD [68, 69]. Zotarolimus exhibits enhanced lipophilicity compared to sirolimus, allowing it to traverse cellular membranes more easily than the less lipophilic drugs sirolimus and paclitaxel [70]. As in sirolimus and everolimus, zotarolimus inhibits smooth muscle cell proliferation via mTOR inhibition [71]. A study assessing reendothelialization rates in an ilio-femoral atherosclerotic rabbit model treated with zotarolimus-compared to everolimus-eluting stents found decreased inflammation and increased expression of CD31, a marker of mature endothelial cells, in the everolimus-treated group compared to the zotarolimus-treated group, suggesting that reendothelialization may occur faster with everolimus than zotarolimus [72]. However, an *in vitro* study examining reendothelialization after inducing an injury in cultured endothelial cells found that regrowth of the injured area occurred more quickly with zotarolimus compared to sirolimus or paclitaxel [73]. While zotarolimus eluting stents have been used in CAD, this technology is not FDA approved for peripheral arteries. Similarly, zotarolimus-coated balloons have been evaluated in pre-clinical studies using the swine femoral artery, however there are no zotarolimus-coated balloons that are FDA approved for treating PAD lesions [74].

Coating technologies and their contributions to efficacy and/or complications

Contributions to efficacy

Unlike DES, which remain in the vessel indefinitely, DCB must be designed such that the coating adheres to the balloon during tracking while also effectively releasing all of their drug/



carrier cargo to the target lesion within a window of 1–3 min. Therefore, the development of effective transfer mechanisms is emphasized in DCB development. Additionally, the coating should have a uniform density and elicit minimal inflammatory responses. The coating should also enhance drug transfer from the device to target tissues with effective tissue penetration and appropriate tissue residence time. Coatings often include excipients, which are defined as ingredients other than the active drug within a formulation. Commonly used excipients in DCBs include urea, polyethylene glycol (PEG), polysorbate and sorbitol, iopromide, and others [75–78]. Excipients can improve drug stability within the vessel environment and modify tissue uptake. Excipients can also act as delivery vehicles, directly transporting drugs to target sites [79–81].

Additional considerations are the hydrophilicity or hydrophobicity of the coating. Hydrophobic coatings create a repellent surface that allows blood to pass over the device. However, hydrophobic coatings may be less hemocompatible than hydrophilic coatings. Multiple studies have associated complement component C3 and fibronectin adsorption as well as monocyte and platelet adhesion with hydrophobic coatings [82–84]. On the other hand, recent investigations into superhydrophobic coatings demonstrate enhanced hemocompatibility. Experimental studies assessing the hemocompatibility of superhydrophobic coatings show reduced protein adsorption and decreased platelet adhesion [85–87].

Polymeric coatings have been used extensively on medical devices. Polymeric hydrophilic coatings exhibit less blood protein adsorption than polymeric hydrophobic coatings, demonstrating enhanced hemocompatibility [82]. Polymeric hydrophilic coatings attract water and tend to be more slippery than hydrophobic coatings, which promotes navigation through tortuous arterial segments during PTA procedures. Additionally, hydrophilic coatings have been shown to

promote rapid drug transfer [88]. In DCBs where the drug load needs to be transferred rapidly from the balloon surface to the treatment site, hydrophilic coatings are advantageous. The downside of hydrophilic coatings is that while they promote drug transfer, they are also prone to significant drug loss from the coating surface during catheter tracking [89, 90].

Aside from the physical properties of the coating, the coating method itself is also an important factor that contributes to efficacy. Several methods exist for applying coatings, each with their own benefits and drawbacks. There are many methods including dip coating, air or ultrasonic spray coating, and others. A study conducted by Gandhi and Murthy found that dip coating balloon catheters created a generally smooth surface, but the coating accumulated around pleated regions of the uninflated balloons, which could alter drug release. The same study showed that an ultrasonic spray coating created a balloon surface with microcracks, while airbrushing created the most uniform surface [91]. Ideal properties of coating and drugs used for DCB are summarized below Figure 2.

Coating distribution, dose, composition and their relations to complications

The interactions between active drugs and the coating are critical components of not only drug delivery, but also the biocompatibility of combination devices. While various excipient coatings have markedly improved drug delivery, coating embolization is also a significant concern in endovascular devices. A study performed by Torii and colleagues compared DCB coating embolization in swine arteries using the IN. PACT, Ranger, and Stellarex DCBs. The IN. PACT DCB utilizes a highly hydrophilic urea coating. In contrast, the Ranger DCB uses an acetyl tributyl citrate carrier that is highly hydrophobic. The study concluded that downstream emboli were found more frequently in the IN. PACT DCB treated arteries [92]. While correlations were drawn between increasing paclitaxel concentration and increased incidence of emboli, it is also possible that the hydrophilicity of the IN. PACT DCB urea coating contributed to embolization via loss during tracking or failure to adhere to the vessel wall due to enhanced affinity for the hydrophilic blood compartment compared to hydrophobic cellular membranes [93]. Previous work in porcine models demonstrated distal excipient-crystalline drug emboli present in the coronary band of pigs after femoral artery angioplasty with paclitaxel-coated balloons [94].

Biodegradable polymers were designed to mitigate inflammatory and thrombotic responses associated with early durable stent coatings. However, biodegradable polymers have also been associated with biocompatibility issues. In the multi-center study conducted by van der Giessen and coworkers, coating materials consisting of both durable and

biodegradable polymers were applied to stents that were implanted into pig arteries and left in place for 4 weeks. All of the polymers tested, including biodegradable PLGA, elicited inflammation in the implanted arteries and lumen narrowing was observed [95]. However, the stents themselves could have contributed to the elicited reaction, as the materials were not sterilized prior to implantation. They also observed that the applied polymer coatings did not fully cover the stents after expansion, leaving areas of bare metal in direct apposition to the treated arteries [95]. Other cited drawbacks associated with PLGA are bulk polymer erosion and burst drug release [96]. Additionally, accumulation of glycolic and lactic acid at local sites due to rapid breakdown of PLGA may incite inflammatory responses [97]. Meanwhile, other studies have found PLGA coated stents to be no more inflammatory than BMS [98]. While PLGA is one of the most widely used polymers in medical devices, continued investigations into PLGA biocompatibility are warranted.

Submicron drug coatings including polymeric nanoparticles are being developed for treatment of PAD lesions [32, 99]. While these technologies have been studied in the setting of cancer treatment and have shown promising results, there are points to consider with the use of submicron particles. Adherence of nanoparticles to the balloon surface during delivery is key; otherwise, there is potential for blood flow to cause early deployment of particles to non-target areas. Blood interactions with nanoparticles are important considerations. Nanoparticle composition, size, and charge are determining factors for how the particles interact with cells in the blood compartment. For example, carbon nanoparticles have been shown to promote venous thrombosis and platelet aggregation [100]. Additionally, some nanoparticles can enter cells such as RBCs by direct penetration of the cell membrane [101]. While carbon nanoparticles have been shown to promote thrombosis, other studies have documented a lack of increased platelet aggregation with the use of PLGA nanoparticles, highlighting the importance of particle type selection for use in blood contacting devices [102]. Bakhaidar and coworkers demonstrated that PLGA-PEG nanoparticles ranging from 112 to 576 nm interacted with and bound to platelets; however, this did not increase platelet aggregation [103].

Emerging devices in BTK PAD therapy

Emerging drug coated balloons

While there have been several clinical trials investigating DCBs for use in BTK PAD lesions, none have emerged as FDA-approved. DCB treatment has not yet demonstrated long term benefits over POBA treatment alone in clinical trials. However, several new DCB have been granted breakthrough device designation and are currently in clinical trials. These newer DCB formulations range from

microcrystalline polymer-free coatings to liquid delivery systems. These combination devices deliver sirolimus as well as combinations of sirolimus and paclitaxel. A table summarizing these novel devices, what we know about their coating properties, and selected clinical trials is listed below [Table 3](#).

The MagicTouch DCB is used in coronary applications in Europe and Asia; however, it has yet to gain FDA approval in the United States. Recently, Concept Medical's MagicTouch DCB received FDA breakthrough designation for BTK PAD lesions. This DCB utilizes a polymer-free approach with Nanolute technology, proprietary 100–300 nm phospholipid microspheres carrying sirolimus. The coating density of the MagicTouch DCB is 1.27 $\mu\text{g}/\text{mm}^2$ [104]. Three-year results from the XTOSI pilot study published in 2024 demonstrate 77.8% freedom from major amputation for BTK lesions [105]. Several clinical trials are currently investigating the use of MagicTouch DCB in both FP and BTK PAD.

Like the MagicTouch DCB, the Sundance™ DCB by SurModics utilizes a polymer-free formulation. The Sundance formulation is a microcrystalline sirolimus coating with their coating density and proprietary excipient yet to be disclosed. In 2020, they commenced the SWING study, a prospective multi-center single arm study that enrolled 35 patients. The completion date for the study was 30 January 2024. SurModics has yet to publish the results of their study, although they reported 71.4% primary patency maintained at 24 months [106].

The Soluton SLR DCB uses sirolimus-loaded PLGA microspheres contained in MicroReservoirs which are coated with Cell Adherent Technology (CAT), a mixture of phospholipids that reportedly protect the microspheres during catheter insertion and tracking. The coating density of the balloon is 1 $\mu\text{g}/\text{mm}^2$ [107]. The PRESTIGE pilot study investigated the performance of the Soluton SLR DCB in occlusive tibial disease and showed 81.5% tibial patency at 6 months [108]. In 2023, the prospective, randomized multicenter single blinded study, SELUTION4BTK completed enrollment with 377 subjects. The aim of the study is to assess the safety and effectiveness of the Soluton SLR DCB in treating BTK PAD in patients with CTLL. The anticipated completion date is 30 July 2029 [109]. In May 2025, investigators reported 12-month data from SUCCESS PTA study at the 2025 New Cardiovascular Horizons meeting. SUCCESS PTA is a single arm post-market surveillance study conducted out of treatment centers in Europe. They reported 2.2% target limb amputation and greater than 90% freedom from clinically driven target lesion revascularization in the 12 months cohort, with an average lesion length of 12–13 cm [110].

While the use of paclitaxel in DCB products is generally being replaced by sirolimus in new generation products, the SirPlux Duo by Advanced NanoTherapies combines both sirolimus and paclitaxel in a novel dual-agent formulation. Paclitaxel and sirolimus are co-encapsulated at a 1:9 w/w ratio within nanoparticles [32]. The coating density, excipient, and composition of the nanoparticle carrier are unclear, although a patent submitted by Advanced NanoTherapies in 2023 suggests

TABLE 3 Recent DCBs with FDA breakthrough designation for BTK PAD and associated clinical trials.

Device	Drug	Coating	Clinical trials
Sundance™ DCB, surmodics inc	Microcrystalline sirolimus <i>Coating density undisclosed</i>	<i>Undisclosed</i>	SWING
Selution™ SLR, cordis	Sirolimus 1 µg/mm ²	PLGA (poly (lactic-co-glycolic acid)) microspheres	PRESTIGE PRISTINE SUCCESS SELUTION4BTK
MagicTouch™ DCB, concept medical	Sirolimus 1.27 µg/mm ²	Sub-micron phospholipid carrier	XTOSI FUTURE-BTK LIMES MAGICAL BTK SIRONA
SirPlux duo Advanced NanoTherapies	Co-encapsulated 1:9 paclitaxel:sirolimus w/w <i>Coating density undisclosed</i>	Nanoparticle, carrier composition undisclosed	ADVANCE-DCB

TABLE 4 BTK trial data for DCB with FDA breakthrough designation status.

Device	Clinical trial	Trial type	Primary patency	CD-TLR (or freedom from CD-TLR)	Freedom from major amputation
Sundance™ DCB, surmodics inc.	SWING [113] (12 months results)	Single arm feasibility study	80%	8%	Not reported
	SWING [106] (24 months results)	Single arm feasibility study	71.4%	8.3%	Not reported
Selution™ SLR, cordis	PRESTIGE [108] (12 months results)	Single arm pilot study	78%	93% freedom from CD-TLR	87% at 12 months
	PRISTINE [114] (12 months results)	Single arm registry study	59.5%	7.4%	72.6% (amputation free survival)
MagicTouch™ DCB, concept medical	XTOSI [105] (36 months results)	Single arm pilot study	50% (at 24 months)	77.8% freedom from CD-TLR	81% at 36 months
Pending trials, results not yet reported:					
Selution™ SLR, cordis	SELUTION4BTK [115]	Randomized controlled trial vs. POBA			
MagicTouch™ DCB, concept medical	FUTURE-BTK [116]	Randomized controlled trial vs. POBA			
	LIMES [117]	Randomized controlled trial vs. POBA			
	MAGICAL BTK [118]	Randomized controlled trial vs. POBA			

that PLGA may be used to entrap paclitaxel and sirolimus [111]. Preclinical work in porcine coronary and femoral arteries and rabbit iliac arteries demonstrate reduced VSMC proliferation with SirPlux Duo compared to paclitaxel DCB treatment [32]. They also investigated particle embolism in a porcine coronary artery model and found a significant reduction in embolized material with the SirPlux Duo DCB. A first-in-human clinical trial investigating the use of SirPlux Duo in patients with *de novo* CAD lesions is currently ongoing [112]. Listed below are some of the early results and up-and-coming clinical trials for DCBs with breakthrough designation status for BTK disease Table 4.

Other balloon-based therapies and novel lesion preparation devices

Aqueous delivery systems that circumvent concerns surrounding the use of drug-coated surfaces in the blood compartment have also been investigated. Atigh et al., delivered paclitaxel via liquid delivery in saline with iohexol as the excipient in *ex vivo* porcine carotid arteries using the Occlusion Perfusion Catheter system by Advanced Catheter Therapies [119]. They were able to demonstrate that the Occlusion Perfusion Catheter effectively delivered the liquid drug into the arterial wall. Similarly, the Virtue SAB by Orchestra BioMed uses a novel AngioInfusion balloon that delivers lyophilized submicron sirolimus in a polyester

nanoparticle carrier via aqueous delivery. This device received breakthrough designation status for BTK PAD, and most recently, investigational device exemption by the FDA in May 2025. Currently, Orchestra Biomed plans to start the VIRTUE trial, a pivotal clinical trial looking at coronary in-stent restenosis.

While not strictly a balloon device, the novel Spur Retrievable Stent System by Reflow Medical has potential to enhance DCB therapy. This device consists of a self-expanding, balloon-delivered stent covered in radial spikes which penetrate the arterial wall. By creating channels in the arterial wall, the Spur device disrupts calcification and enhances vessel compliance, reducing elastic recoil immediately after treatment [120]. Additionally, these arterial channels may increase DCB drug penetration. Previously completed clinical trials investigating the use of the Spur BTK followed by DCB therapy include the DEEPER [121], DEEPER OUS [120], and most recently, the DEEPER LIMUS [122] trials.

Stents and resorbable scaffolds

While this review focuses primarily on balloon-based therapies, we would be remiss to exclude these devices from discussion. While the “leave nothing behind” approach has favored DCB for BTK disease, stents and resorbable scaffolds have been gaining traction. Several stents and resorbable scaffolds have gained FDA breakthrough designation status, and one resorbable scaffold, the Esprit BTK everolimus-eluting bioresorbable scaffold by Abbott, recently gained FDA approval [123].

Resorbable scaffolds aim to bridge the gap between providing structural support and minimizing permanent implants. In April 2024, Abbott’s Esprit BTK everolimus-eluting bioresorbable scaffold received FDA approval [123] for CTLI in BTK disease. According to Abbott, this bioresorbable stent maintains radial strength similar to metal stents for the first 6 months and fully dissolves within two to 3 years. In the LIFE-BTK trial, Abbott reported substantially improved efficacy compared to POBA with respect to primary efficacy endpoints at 1 year [124]. Two-year follow up demonstrated continued superior efficacy over POBA, with respect to a composite of limb salvage and primary patency [125].

In March 2024, the Biotronik Freesolve BTK RMS received breakthrough designation status for CTLI BTK disease. This resorbable metal scaffold consists of a proprietary magnesium alloy utilizing sirolimus to treat lesioned arteries [126]. The first-in-human BIOMAG I trial examined late lumen loss (LLL) as a primary endpoint in coronary artery disease. Between 6 and 12 months, significant increase in in-device LLL was reported, however no scaffold thrombosis was observed [126]. The BIOMAG II RCT trial began enrollment in May 2024, and will compare safety and efficacy endpoints versus the Xience everolimus-eluting stent [127].

Efemoral medical’s Efemoral Vascular Scaffold System also received breakthrough designation status in 2024. This

bioresorbable system, like the Freesolve, utilizes sirolimus as an antirestenotic agent. The Efemoral vascular scaffold system focuses on enhancing biomechanical compatibility, utilizing a patented FlexStep system which utilizes interscaffold spaces to enhance device flexibility [128]. The first-in-human EFEMORAL I trial is still ongoing (NCT: 04584632).

Another notable stent to receive FDA breakthrough designation status for BTK indications includes Elixir Medical’s DynamX BTK system. The DynamX BTK Bioadaptor represents a hybrid between bioresorbable polymer elements and metallic scaffolding materials which deploy as the polymer dissolves [129]. The bioresorbable coating consists of poly-L-lactide (PLLA) eluting novolimus [129] and is currently undergoing clinical trials for coronary applications, with plans to design a modified version of the device for BTK therapy [130].

Discussion

PAD is a complex and multifactorial disease that leaves patients prone to thrombosis, arterial occlusion, and loss of limb. Endothelial injury and dysfunction play a key role in the pathogenesis of PAD. PAD below the knee is challenging to treat due to the types of lesions, extent of lesions, as well as significant problems with calcification as a barrier to drug penetration and vascular elastic recoil limiting luminal diameter post-treatment. Yet, numerous novel devices are on the horizon. While DCBs for use below the knee have yet to gain FDA approval, lessons learned from previous device iterations pave the way forward for next-generation devices. Further advancement in coating technology and drug delivery systems permit the use of less drug than older generation devices and rely less on large particulate and crystalline coatings. These changes may limit toxicity off target associated with commonly used drugs like paclitaxel and sirolimus. Drug eluting stents and bioresorbable scaffolds have been gaining momentum in BTK disease treatment and provide another promising avenue for interventions. Abbott’s Esprit BTK received FDA approval, and several bioresorbable scaffolds and DES received FDA breakthrough designation status in just the past year. As clinical trials progress, we will discover whether these breakthrough therapies can gain FDA approval for BTK disease treatment.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Funding

The authors declare that financial support was received for the research and/or publication of this article. Funds for this

work were provided through discretionary funding through the LSU School of Veterinary Medicine (PM20GM130555).

Conflict of interest

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

References

- Criqui MH, Matsushita K, Aboyans V, Hess CN, Hicks CW, Kwan TW, et al. Lower extremity peripheral artery disease: contemporary epidemiology, management gaps, and future directions: a scientific statement from the American heart association. *Circulation* (2021) **144**:e171–e191. doi:10.1161/cir.0000000000001005
- Agnelli G, Belch JJF, Baumgartner I, Giovass P, Hoffmann U. Morbidity and mortality associated with atherosclerotic peripheral artery disease: a systematic review. *Atherosclerosis* (2020) **293**:94–100. doi:10.1016/j.atherosclerosis.2019.09.012
- Bevan GH, White Solaru KT. Evidence-based medical management of peripheral artery disease. *Arteriosclerosis, Thromb Vasc Biol* (2020) **40**:541–53. doi:10.1161/atvbaha.119.312142
- Gornik HL, Aronow HD, Goodney PP, Arya S, Brewster LP, Byrd L, et al. 2024 ACC/AHA/AACVPR/APMA/ABC/SCAI/SVM/SVN/SVS/SIR/VES guideline for the management of lower extremity peripheral artery disease: a report of the American college of cardiology/american heart association joint committee on clinical practice guidelines. *Circulation* (2024) **149**:e1313–e1410. doi:10.1161/CIR.0000000000001251
- Thukkani AK, Kinlay S. Endovascular intervention for peripheral artery disease. *Circ Res* (2015) **116**:1599–613. doi:10.1161/circresaha.116.303503
- Liistro F, Angioli P, Venturuzzo G, Ducci K, Reccia MR, Ricci L, et al. Randomized controlled trial of acotec drug-eluting balloon versus plain balloon for below-the-knee angioplasty. *JACC: Cardiovasc Interventions* (2020) **13**:2277–86. doi:10.1016/j.jcin.2020.06.045
- Narula N, Dannenberg AJ, Olin JW, Bhatt DL, Johnson KW, Nadkarni G, et al. Pathology of peripheral artery disease in patients with critical limb ischemia. *J Am Coll Cardiol* (2018) **72**:2152–63. doi:10.1016/j.jacc.2018.08.002
- Smolenski A. Novel roles of cAMP/cGMP dependent signaling in platelets. *J Thromb Haemost* (2012) **10**:167–76. doi:10.1111/j.1538-7836.2011.04576.x
- Wood JP, Bunce MW, Maroney SA, Tracy PB, Camire RM, Mast AE. Tissue factor pathway inhibitor-alpha inhibits prothrombinase during the initiation of blood coagulation. *Proc Natl Acad Sci U S A* (2013) **110**:17838–43. doi:10.1073/pnas.1310444110
- Rhee M, Lee J, Lee EY, Yoon K-H, Lee S-H. Lipid variability induces endothelial dysfunction by increasing inflammation and oxidative stress. *Endocrinol Metab* (2024) **39**:511–20. doi:10.3803/enm.2023.1915
- Sellak H, Franzini E, Hakim J, Pasquier C. Reactive oxygen species rapidly increase endothelial ICAM-1 ability to bind neutrophils without detectable upregulation. *Blood* (1994) **83**:2669–77. doi:10.1182/blood.v83.9.2669.bloodjournal8392669
- De Caterina R, Libby P, Peng HB, Thannickal VJ, Rajavashisth TB, Gimbrone MA, et al. Nitric oxide decreases cytokine-induced endothelial activation. Nitric oxide selectively reduces endothelial expression of adhesion molecules and proinflammatory cytokines. *J Clin Invest* (1995) **96**:60–8. doi:10.1172/jci118074
- Kubes P, Suzuki M, Granger DN. Nitric oxide: an endogenous modulator of leukocyte adhesion. *Proc Natl Acad Sci* (1991) **88**:4651–5. doi:10.1073/pnas.88.11.4651
- Badimon L, Vilahur G. Thrombosis formation on atherosclerotic lesions and plaque rupture. *J Intern Med* (2014) **276**:618–32. doi:10.1111/joim.12296
- Rafieian-Kopaei M, Setorki M, Douadi M, Baradaran A, Nasri H. Atherosclerosis: process, indicators, risk factors and new hopes. *Int J Prev Med* (2014) **5**:927–46.
- Fernández-Ortiz A, Badimon JJ, Falk E, Fuster V, Meyer B, Mailhac A, et al. Characterization of the relative thrombogenicity of atherosclerotic plaque

Generative AI statement

The authors declare that no Generative AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

components: implications for consequences of plaque rupture. *J Am Coll Cardiol* (1994) **23**:1562–9. doi:10.1016/0735-1097(94)90657-2

17. Torii S, Mustapha JA, Narula J, Mori H, Saab F, Jinnouchi H, et al. Histopathologic characterization of peripheral arteries in subjects with abundant risk factors: correlating imaging with pathology. *JACC: Cardiovasc Imaging* (2019) **12**:1501–13. doi:10.1016/j.jcmg.2018.08.039

18. Chang Z, Yan H, Zhen Y, Zheng J, Liu Z. Lower limb arterial calcification and acute thrombosis risk in patients with peripheral artery disease. *Ann Vasc Surg* (2020) **63**:227–33. doi:10.1016/j.avsg.2019.06.043

19. Narula N, Olin JW, Narula N. Pathologic disparities between peripheral artery disease and coronary artery disease. *Arteriosclerosis, Thromb Vasc Biol* (2020) **40**:1982–9. doi:10.1161/atvbaha.119.312864

20. Konijn LCD, Takx RAP, Mali WPTM, Veger HTC, van Overhagen H. Different lower extremity arterial calcification patterns in patients with chronic limb-threatening ischemia compared with asymptomatic controls. *J Pers Med* (2021) **11**:493. doi:10.3390/jpm11060493

21. Konijn LCD, Takx RAP, Jong PA, Spreen MI, Veger HTC, Mali WPTM, et al. Arterial calcification and long-term outcome in chronic limb-threatening ischemia patients. *Eur J Radiol* (2025) **132**. doi:10.1016/j.ejrad.2020.109305

22. Block PC, Myler RK, Stertzer S, Fallon JT. Morphology after transluminal angioplasty in human beings. *New Engl J Med* (1981) **305**:382–5. doi:10.1056/nejm198108133050706

23. Bryckaert M, Rosa J-P, Denis CV, Lenting PJ. Of von Willebrand factor and platelets. *Cell Mol Life Sci* (2014) **72**:307–26. doi:10.1007/s00018-014-1743-8

24. Daniel JL, Dangelmaier C, Jin J, Ashby B, Smith JB, Kunapuli SP. Molecular basis for ADP-Induced platelet activation: I. Evidence for three distinct adp receptors on human platelets. *J Biol Chem* (1998) **273**:2024–9. doi:10.1074/jbc.273.4.2024

25. Nakagaki T, Foster DC, Berkner KL, Kiesel W. Initiation of the extrinsic pathway of blood coagulation: evidence for the tissue factor dependent autoactivation of human coagulation factor VII. *Biochemistry* (1991) **30**:10819–24. doi:10.1021/bi00109a001

26. Ha JM, Yun SJ, Kim YW, Jin SY, Lee HS, Song SH, et al. Platelet-derived growth factor regulates vascular smooth muscle phenotype via mammalian target of rapamycin complex 1. *Biochem Biophysical Res Commun* (2015) **464**:57–62. doi:10.1016/j.bbrc.2015.05.097

27. Sato Y, Hamanaka R, Ono J, Kuwano M, Rifkin DB, Takaki R. The stimulatory effect of PDGF on vascular smooth muscle cell migration is mediated by the induction of endogenous basic FGF. *Biochem Biophysical Res Commun* (1991) **174**:1260–6. doi:10.1016/0006-291x(91)91557-s

28. Kohno K, Koya-Miyata S, Harashima A, Tsukuda T, Katakami M, Ariyasu T, et al. Inflammatory M1-like macrophages polarized by NK-4 undergo enhanced phenotypic switching to an anti-inflammatory M2-like phenotype upon co-culture with apoptotic cells. *J Inflamm* (2021) **18**:2. doi:10.1186/s12950-020-00267-z

29. Choi S, Park M, Kim J, Park W, Kim S, Lee D-K, et al. TNF- α elicits phenotypic and functional alterations of vascular smooth muscle cells by miR-155-5p-dependent down-regulation of cGMP-dependent kinase 1. *J Biol Chem* (2018) **293**:14812–22. doi:10.1074/jbc.ra118.004220

30. Jovinge S, Hultgardrdh-Nilsson A, Regnström J, Nilsson J. Tumor necrosis Factor- α activates smooth muscle cell migration in culture and is expressed in the balloon-injured rat aorta. *Arteriosclerosis, Thromb Vasc Biol* (1997) **17**:490–7. doi:10.1161/01.atv.17.3.490

31. Rectenwald JE, Moldawer LL, Huber TS, Seeger JM, Ozaki CK. Direct evidence for cytokine involvement in neointimal hyperplasia. *Circulation* (2000) **102**:1697–702. doi:10.1161/01.cir.102.14.1697

32. Kawai K, Rahman MT, Nowicki R, Kolodgie FD, Sakamoto A, Kawakami R, et al. Efficacy and safety of dual paclitaxel and sirolimus nanoparticle-coated balloon. *JACC: Basic Translational Sci* (2024) **9**:774–89. doi:10.1016/j.jacpts.2024.02.002
33. Shishebor MH, Scheinert D, Jain A, Brodmann M, Tepe G, Ando K, et al. Comparison of drug-coated balloons vs bare-metal stents in patients with femoropopliteal arterial disease. *J Am Coll Cardiol* (2023) **81**:237–49. doi:10.1016/j.jacc.2022.10.016
34. Beckman JA, Schneider PA, Conte MS. Advances in revascularization for peripheral artery disease: revascularization in PAD. *Circ Res* (2021) **128**:1885–912. doi:10.1161/circresaha.121.318261
35. Axel DI, Kunert W, Göggelmann C, Oberhoff M, Herdeg C, Küttner A, et al. Paclitaxel inhibits arterial smooth muscle cell proliferation and migration *in vitro* and *in vivo* using local drug delivery. *Circulation* (1997) **96**:636–45. doi:10.1161/01.cir.96.2.636
36. Grube E, Silber S, Hauptmann KE, Mueller R, Buellesfeld L, Gerckens U, et al. TAXUS I: six- and twelve-month results from a randomized, double-blind trial on a slow-release paclitaxel-eluting stent for *de novo* coronary lesions. *Circulation* (2003) **107**:38–42. doi:10.1161/01.cir.0000047700.58683.a1
37. Unverdorben M, Kleber FX, Heuer H, Figulla H-R, Vallbracht C, Leschke M, et al. Treatment of small coronary arteries with a paclitaxel-coated balloon catheter. *Clin Res Cardiol* (2010) **99**:165–74. doi:10.1007/s00392-009-0101-6
38. Rosenfield K, Jaff MR, White CJ, Rocha-Singh K, Mena-Hurtado C, Metzger DC, et al. Trial of a paclitaxel-coated balloon for femoropopliteal artery disease. *New Engl J Med* (2015) **373**:145–53. doi:10.1056/nejmoa1406235
39. Shu CH, Yang WK, Shih YL, Kuo ML, Huang TS. Cell cycle G2/M arrest and activation of cyclin-dependent kinases associated with low-dose paclitaxel-induced sub-G1 apoptosis. *Apoptosis* (1997) **2**:463–70. doi:10.1023/a:1026422111457
40. Nguyen KT, Shaikh N, Wawro D, Zhang S, Schwade ND, Eberhart RC, et al. Molecular responses of vascular smooth muscle cells to paclitaxel-eluting bioresorbable stent materials. *J Biomed Mater Res A* (2004) **69A**:513–24. doi:10.1002/jbm.a.30020
41. Nakazawa G, Finn AV, Virmani R. Vascular pathology of drug-eluting stents. *Herz* (2007) **32**:274–80. doi:10.1007/s00059-007-2997-9
42. Katsanos K, Spiliopoulos S, Kitrou P, Krokidis M, Karnabatidis D. Risk of death following application of paclitaxel-coated balloons and stents in the femoropopliteal artery of the leg: a systematic review and meta-analysis of randomized controlled trials. *J Am Heart Assoc* (2018) **7**:e011245. doi:10.1161/jaha.118.011245
43. Bittl JA, He Y, Baber U, Feldman RL, von Mering GO, Kaul S. Bayes factor meta-analysis of the mortality claim for peripheral paclitaxel-eluting devices. *JACC: Cardiovasc Interventions* (2019) **12**:2528–37. doi:10.1016/j.jcin.2019.09.028
44. Rocha-Singh KJ, Duval S, Jaff MR, Schneider PA, Ansel GM, Lyden SP, et al. Mortality and paclitaxel-coated devices: an individual patient data meta-analysis. *Circulation* (2020) **141**:1859–69. doi:10.1161/circulationaha.119.044697
45. Dinh K, Limmer AM, Chen AZL, Thomas SD, Holden A, Schneider PA, et al. Mortality rates after paclitaxel-coated device use in patients with occlusive femoropopliteal disease: an updated systematic review and meta-analysis of randomized controlled trials. *J Endovasc Ther* (2021) **28**:755–77. doi:10.1177/15266028211023505
46. Health CD. UPDATE: paclitaxel-coated devices to treat peripheral arterial disease unlikely to increase risk of mortality - letter to health care providers. *FDA* (2023). Available online at: <https://www.fda.gov/medical-devices/letters-health-care-providers/update-paclitaxel-coated-devices-treat-peripheral-arterial-disease-unlikely-increase-risk-mortality> (Accessed July 9, 2025).
47. Sehgal SN. Sirolimus: its discovery, biological properties, and mechanism of action. *Transplant Proc* (2003) **35**:7S–14S. doi:10.1016/s0041-1345(03)00211-2
48. Liu Y, Li X, Jin A. Rapamycin inhibits Nf-KB activation by autophagy to reduce catabolism in human chondrocytes. *J Invest Surg* (2020) **33**:861–73. doi:10.1080/08941939.2019.1574321
49. Lee J-J, Yu J-Y, Lee J-H, Zhang WY, Kim T-J, Myung C-S, et al. The protective effects of paclitaxel on platelet aggregation through the inhibition of thromboxane A2 synthase. *Arch Pharm Res* (2010) **33**:387–94. doi:10.1007/s12272-010-0307-1
50. Zhang S, Sun C, Hu H, He Y, Yao Y, Cao Y, et al. Effects of paclitaxel on the ability of aspirin and clopidogrel to inhibit platelet aggregation. *Clin Appl Thromb Hemost* (2016) **22**:673–8. doi:10.1177/1076029615576740
51. Wu Q, Huang K-S, Chen M, Huang D-J. Rapamycin enhances platelet aggregation induced by adenosine diphosphate *in vitro*. *Platelets* (2009) **20**:428–31. doi:10.1080/09537100903114552
52. Babinska A, Markell MS, Salifu MO, Akoad M, Ehrlich YH, Kornecki E. Enhancement of human platelet aggregation and secretion induced by rapamycin. *Nephrol Dial Transplant* (1998) **13**:3153–9. doi:10.1093/ndt/13.12.3153
53. Zhang M, Lotfollahzadeh S, Elzain N, Yang X, Elsadawi M, Gower AC, et al. Alleviating iatrogenic effects of paclitaxel *via* antiinflammatory treatment. *Vasc Med* (2024) **29**:369–80. doi:10.1177/1358863x241231942
54. Farb A, Heller PF, Shroff S, Cheng L, Kolodgie FD, Carter AJ, et al. Pathological analysis of local delivery of paclitaxel *via* a polymer-coated stent. *Circulation* (2001) **104**:473–9. doi:10.1161/hc3001.092037
55. Nührenberg TG, Voisard R, Fahlisch F, Rudelius M, Braun J, Gschwend J, et al. Rapamycin attenuates vascular wall inflammation and progenitor cell promoters after angioplasty. *The FASEB J* (2005) **19**:1–21. doi:10.1096/fj.04.2431fje
56. Kahan BD, Chang JY, Sehgal SN. Preclinical evaluation of a new potent immunosuppressive agent, rapamycin. *Transplantation* (1991) **52**:185–91. doi:10.1097/00007890-199108000-00001
57. Marx SO, Jayaraman T, Go LO, Marks AR. Rapamycin-FKBP inhibits cell cycle regulators of proliferation in vascular smooth muscle cells. *Circ Res* (1995) **76**:412–7. doi:10.1161/01.res.76.3.412
58. Poon M, Marx SO, Gallo R, Badimon JJ, Taubman MB, Marks AR. Rapamycin inhibits vascular smooth muscle cell migration. *J Clin Invest* (1996) **98**:2277–83. doi:10.1172/jci119038
59. Khing TM, Choi WS, Kim DM, Po WW, Thein W, Shin CY, et al. The effect of paclitaxel on apoptosis, autophagy and mitotic catastrophe in AGS cells. *Sci Rep* (2021) **11**:23490. doi:10.1038/s41598-021-02503-9
60. Belotti D, Vergani V, Drudis T, Borsotti P, Pitelli MR, Viale G, et al. The microtubule-affecting drug paclitaxel has antiangiogenic activity. *Clin Cancer Res* (1996) **2**:1843–9.
61. Liu H-T, Li F, Wang W-Y, Li X-J, Liu Y-M, Wang R-A, et al. Rapamycin inhibits Re-Endothelialization after percutaneous coronary intervention by impeding the proliferation and migration of endothelial cells and inducing apoptosis of endothelial progenitor cells. *Tex Heart Inst J* (2010) **37**:194–201.
62. Sedrani R, Cottens S, Kallen J, Schuler W. Chemical modification of rapamycin: the discovery of SDZ RAD. *Transplant Proc* (1998) **30**:2192–4. doi:10.1016/s0041-1345(98)00587-9
63. Schuler W, Sedrani R, Cottens S, Häberlin B, Schulz M, Schuurman HJ, et al. SDZ RAD, a new rapamycin derivative: pharmacological properties *in vitro* and *in vivo*. *Transplantation* (1997) **64**:36–42. doi:10.1097/00007890-199707150-00008
64. Schreiber KH, Ortiz D, Academia EC, Anies AC, Liao C-Y, Kennedy BK. Rapamycin-mediated mTORC2 inhibition is determined by the relative expression of FK506-binding proteins. *Aging Cell* (2015) **14**:265–73. doi:10.1111/accel.12313
65. Chen G, Ding X-F, Bouamar H, Pressley K, Sun L-Z. Everolimus induces G1 cell cycle arrest through autophagy-mediated protein degradation of cyclin D1 in breast cancer cells. *Am J Physiology-Cell Physiol* (2019) **317**:C244–C252. doi:10.1152/ajpcell.00390.2018
66. Tsuchida K, Piek JJ, Neumann F-J, van der Giessen WJ, Wiemer M, Zeiher AM, et al. One-year results of a durable polymer everolimus-eluting stent in *de novo* coronary narrowings (The SPIRIT FIRST Trial). *EuroIntervention* (2005) **1**:266–72.
67. Grube E, Sonoda S, Ikeno F, Honda Y, Kar S, Chan C, et al. Six- and twelve-month results from first human experience using everolimus-eluting stents with bioabsorbable polymer. *Circulation* (2004) **109**:2168–71. doi:10.1161/01.cir.0000128850.84227.f0
68. Leone PP, Assafin M, Scotti A, Gonzalez M, Mignatti A, Dawson K, et al. A technology evaluation of the onyx frontier drug-eluting stent. *Expert Opin Drug Deliv* (2023) **20**:689–701. doi:10.1080/17425247.2023.2216449
69. Kirtane AJ, Leon MB, Ball MW, Bajwa HS, Sketch MH, Coleman PS, et al. The “final” 5-Year Follow-Up from the ENDEAVOR IV trial comparing a zotarolimus-eluting stent with a paclitaxel-eluting stent. *JACC: Cardiovasc Interventions* (2013) **6**:325–33. doi:10.1016/j.jcin.2012.12.123
70. Burke SE, Kuntz RE, Schwartz LB. Zotarolimus (ABT-578) eluting stents. *Adv Drug Deliv Rev* (2006) **58**:437–46. doi:10.1016/j.addr.2006.01.021
71. Chen Y-W, Smith ML, Sheets M, Ballaron S, Trevillyan JM, Burke SE, et al. Zotarolimus, a novel sirolimus analogue with potent anti-proliferative activity on coronary smooth muscle cells and reduced potential for systemic immunosuppression. *J Cardiovasc Pharmacol* (2007) **49**:228–35. doi:10.1097/fjc.0b013e3180325b0a
72. Yazdani SK, Sheehy A, Nakano M, Nakazawa G, Vorpahl M, Otsuka F, et al. Preclinical evaluation of second-generation Everolimus- and zotarolimus-eluting coronary stents. *The J Invasive Cardiology* (2013) **25**:383–90. Available online at: <https://www.hmpgloblearningnetwork.com/site/jic/articles/preclinical-evaluation-second-generation-everolimus-and-zotarolimus-eluting-coronary-stents> (Accessed August 23, 2024).
73. Miura K, Nakaya H, Kobayashi Y. Experimental assessment of effects of antiproliferative drugs of drug-eluting stents on endothelial cells. *Cardiovasc Revascularization Med* (2015) **16**:344–7. doi:10.1016/j.carrev.2015.07.002

74. Granada JF, Milewski K, Zhao H, Stankus JJ, Tellez A, Aboodi MS, et al. Vascular response to zotarolimus-coated balloons in injured superficial femoral arteries of the familial hypercholesterolemic swine. *Circ Cardiovasc Interventions* (2011) 4:447–55. doi:10.1161/circinterventions.110.960260
75. Sachar R, Soga Y, Ansari MM, Kozuki A, Lopez L, Brodmann M, et al. 1-Year results from the RANGER II SFA randomized trial of the ranger drug-coated balloon. *JACC: Cardiovasc Interventions* (2021) 14:1123–33. doi:10.1016/j.jcin.2021.03.021
76. Gruber P, Braun C, Kahles T, Hlavica M, Anon J, Diepers M, et al. Percutaneous transluminal angioplasty using the novel drug-coated balloon catheter SeQuent please NEO for the treatment of symptomatic intracranial severe stenosis: feasibility and safety study. *J NeuroInterventional Surg* (2019) 11:719–22. doi:10.1136/neurintsurg-2018-014378
77. Chen Z, Guo W, Jiang W, Wang F, Fu W, Zou Y, et al. IN.PACT SFA clinical study using the IN.PACT admiral drug-coated balloon in a Chinese patient population. *J Endovasc Ther* (2019) 26:471–8. doi:10.1177/1526602819852084
78. Krishnan P, Faries P, Niazi K, Jain A, Sachar R, Bachinsky WB, et al. Stellarex drug-coated balloon for treatment of femoropopliteal disease. *Circulation* (2017) 136:1102–13. doi:10.1161/circulationaha.117.028893
79. Chen R, Liu E, Fang Y, Gao N, Zhang M, Zhang X, et al. Naturally sourced amphiphilic peptides as paclitaxel vehicles for breast cancer treatment. *Biomater Adv* (2024) 159:213824. doi:10.1016/j.bioadv.2024.213824
80. Sodha S, Gupta P. PLGA and PEG based porous microparticles as vehicles for pulmonary somatropin delivery. *Eur J Pharmaceutics Biopharmaceutics* (2023) 191:150–7. doi:10.1016/j.ejpb.2023.08.017
81. Codoni D, Cowan J, Bradley J, McAuley WJ, O'Connell MA, Qi S. Disc-shaped polyoxyethylene glycol glycerides gel nanoparticles as novel protein delivery vehicles. *Int J Pharmaceutics* (2015) 496:1015–25. doi:10.1016/j.ijpharm.2015.10.067
82. Hezi-Yamit A, Sullivan C, Wong J, David L, Chen M, Cheng P, et al. Impact of polymer hydrophilicity on biocompatibility: implication for DES polymer design. *J Biomed Mater Res Part A* (2009) 90A:133–41. doi:10.1002/jbm.a.32057
83. Collier TO, Jenney CR, DeFife KM, Anderson JM. Protein adsorption on chemically modified surfaces. *Biomed Sci Instrumentation* (1997) 33:178–83.
84. Yayapour N, Nygren H. Interactions between whole blood and hydrophilic or hydrophobic glass surfaces: kinetics of cell adhesion. *Colloids Surf B: Biointerfaces* (1999) 15:127–38. doi:10.1016/s0927-7765(99)00049-1
85. Zhang W, Du J, Zhu T, Wang R. SiO₂ nanosphere coated tough catheter with superhydrophobic surface for improving the antibacteria and hemocompatibility. *Front Bioeng Biotechnol* (2023) 10:1067139. doi:10.3389/fbioe.2022.1067139
86. Zhang J, Li G, Li D, Zhang X, Li Q, Liu Z, et al. *In vivo* blood-repellent performance of a controllable facile-generated superhydrophobic surface. *ACS Appl Mater Inter* (2021) 13:29021–33. doi:10.1021/acsami.0c21058
87. Movafaghi S, Leszczak V, Wang W, Sorkin JA, Dasi LP, Popat KC, et al. Hemocompatibility of superhydrophobic titania surfaces. *Adv Healthc Mater* (2017) 6:1600717. doi:10.1002/adhm.201600717
88. Shazly T, Eberth JF, Kostelnik CJ, Uline MJ, Chitalia VC, Spinale FG, et al. Hydrophilic coating microstructure mediates acute drug transfer in drug-coated balloon therapy. *ACS Appl Bio Mater* (2024) 7:3041–9. doi:10.1021/acsabm.4c00080
89. Anbalakan K, Toh HW, Ang HY, Buist ML, Leo HL. How does the nature of an excipient and an atheroma influence drug-coated balloon therapy? *Cardiovasc Eng Tech* (2022) 13:915–29. doi:10.1007/s13239-022-00626-2
90. Anderson JA, Remund T, Pohlson K, Lamichane S, Evans C, Evans R, et al. *In vitro* and *in vivo* evaluation of effect of excipients in local delivery of paclitaxel using microporous infusion balloon catheters. *J Biomed Mater Res B: Appl Biomater* (2017) 105:376–90. doi:10.1002/jbm.b.33564
91. Gandhi PJ, Murthy ZVP. Investigation of different drug deposition techniques on drug releasing properties of cardiovascular drug coated balloons. *Ind Eng Chem Res* (2012) 51:10800–23. doi:10.1021/ie3006676
92. Torii S, Jinnouchi H, Sakamoto A, Romero ME, Kolodgie FD, Virmani R, et al. Comparison of biologic effect and particulate embolization after femoral artery treatment with three drug-coated balloons in healthy swine model. *J Vasc Interv Radiol* (2019) 30:103–9. doi:10.1016/j.jvir.2018.07.025
93. Heilmann T, Richter C, Noack H, Post S, Mahnkopf D, Mittag A, et al. Drug release profiles of different drug-coated balloon platforms. *Eur Cardiol Rev* (2010) 6:40. doi:10.15420/ecn.2010.8.2.40
94. Kolodgie FD, Pacheco E, Yahagi K, Mori H, Ladich E, Virmani R. Comparison of particulate embolization after femoral artery treatment with IN.PACT admiral versus lutonix 035 paclitaxel-coated balloons in healthy swine. *J Vasc Interv Radiol* (2016) 27:1676–85.e2. doi:10.1016/j.jvir.2016.06.036
95. van der Giessen WJ, Lincoff AM, Schwartz RS, van Beusekom HMM, Serruys PW, Holmes DR, et al. Marked inflammatory sequelae to implantation of biodegradable and nonbiodegradable polymers in porcine coronary arteries. *Circulation* (1996) 94:1690–7. doi:10.1161/01.cir.94.7.1690
96. Bakhrushina EO, Sakharova PS, Konogorova PD, Pyzhov VS, Kosenkova SI, Bardakov AI, et al. Burst release from *in situ* forming PLGA-based implants: 12 effectors and ways of correction. *Pharmaceutics* (2024) 16:115. doi:10.3390/pharmaceutics16010115
97. Ma S, Feng X, Liu F, Wang B, Zhang H, Niu X. The pro-inflammatory response of macrophages regulated by acid degradation products of poly(lactide-co-glycolide) nanoparticles. *Eng Life Sci* (2021) 21:709–20. doi:10.1002/elsc.202100040
98. Peng H-Y, Chen M, Zheng B, Wang X-G, Huo Y. Long-term effects of novel biodegradable, polymer-coated, Sirolimus-Eluting stents on neointimal formation in a porcine coronary model. *Int Heart J* (2009) 50:811–22. doi:10.1536/ihj.50.811
99. Iida O, Soga Y, Saito S, Mano T, Hayakawa N, Ichihashi S, et al. A novel Sirolimus-Coated balloon for the treatment of femoropopliteal lesions. *JACC: Cardiovasc Interventions* (2024) 17:1547–56. doi:10.1016/j.jcin.2024.03.029
100. Radomski A, Jurasz P, Alonso-Escobedo D, Drews M, Morandi M, Malinski T, et al. Nanoparticle-induced platelet aggregation and vascular thrombosis. *Br J Pharmacol* (2005) 146:882–93. doi:10.1038/sj.bjp.0706386
101. Wang T, Bai J, Jiang X, Nienhaus GU. Cellular uptake of nanoparticles by membrane penetration: a study combining confocal microscopy with FTIR spectroelectrochemistry. *ACS Nano* (2012) 6:1251–9. doi:10.1021/nn203892h
102. Li X, Radomski A, Corrigan OI, Tajber L, De Sousa Menezes F, Endter S, et al. Platelet compatibility of PLGA, chitosan and PLGA–chitosan nanoparticles. *Nanomedicine* (2009) 4:735–46. doi:10.2217/nnm.09.65
103. Bakhaidar R, Green J, Alfahad K, Samanani S, Moollan N, O'Neill S, et al. Effect of size and concentration of PLGA-PEG nanoparticles on activation and aggregation of washed human platelets. *Pharmaceutics* (2019) 11:514. doi:10.3390/pharmaceutics11100514
104. Ninomiya K, Serruys PW, Colombo A, Reimers B, Basavarajiah S, Sharif F, et al. A Prospective Randomized Trial Comparing Sirolimus-Coated Balloon With Paclitaxel-Coated Balloon in *de novo* Small Vessels. *JACC: Cardiovasc Interventions* (2023) 16:2884–96. doi:10.1016/j.jcin.2023.09.026
105. Choke ETC, Peh EYL, Tang TY, Cheng SC, Tay JS, Aw DKL, et al. MagicTouch PTA Sirolimus-Coated balloon for femoropopliteal and below-the-knee disease: 3-year outcomes of the XTOSI trial. *Ann Vasc Surg* (2024) 106:8–15. doi:10.1016/j.avsg.2023.12.096
106. Surmodics announces 24-Month data from the SWING trial presented at VEITHsymposium surmodics. Inc. (2025). Available online at: <https://surmodics.gcs-web.com/news-releases/news-release-details/surmodics-announces-24-month-data-swing-trial-presented> (Accessed 30 January 2025).
107. Tang TY, Chong T-T, Yap CJQ, Soon SXY, Chan SL, Tan RY, et al. Intervention with solution SLR™ agent balloon for endovascular latent limbus therapy for failing AV fistulas (ISABELLA) trial: protocol for a pilot clinical study and pre-clinical results. *J Vasc Access* (2023) 24:289–99. doi:10.1177/11297298211020867
108. Tang TY, Yap C, Soon SXY, Chan SL, Lee QS, Yap HY, et al. World's first experience treating TASC II C and D tibial occlusive disease using the solution SLR Sirolimus-Eluting balloon: six-month results from the PRESTIGE study. *J Endovasc Ther* (2021) 28:555–66. doi:10.1177/15266028211007457
109. Armstrong E. CRT-300.3 SELUTION4BTK – a randomized clinical trial evaluating solution SLR Sirolimus-Eluting balloon in the treatment of below-the-knee lesions in patients with chronic limb-threatening ischemia. *JACC: Cardiovasc Interventions* (2023) 16:S50. doi:10.1016/j.jcin.2023.01.162
110. Ncvh 2025 agenda – NCVH (2025). Available online at: <https://ncvh.org/ncvh-2025-agenda/> (Accessed 10 July 2025).
111. Berrada-Sounni M, Zuckerman ST. Undulating balloon systems and methods for nanoparticle-based drug delivery. *US20230106928A1* (2023). Available online at: <https://patents.google.com/patent/US20230106928A1/de> (Accessed February 3, 2025).
112. NanoTherapies A. ADVANCED NanoTherapies dual active pharmacological ingredient (Dual-API) drug-coated balloon catheter to treat de-novo lesions in patients with symptomatic stable angina, unstable angina, and NSTEMI. Clinical trial registration NCT05521542, clinicaltrials.gov (2025). Available online at: <https://clinicaltrials.gov/study/NCT05521542> (Accessed July 10, 2024).
113. Surmodics announces 12-Month data from the SWING trial presented at VEITHsymposium | surmodics. Inc. (2025) Available online at: <https://surmodics.gcs-web.com/news-releases/news-release-details/surmodics-announces-12-month-data-swing-trial-presented> (Accessed 29 October 2025).
114. Tang TY, Yap C, Chan SL, Soon SXY, Sivanathan C, Gogna A, et al. The utility of sirolimus eluting balloons in the setting of chronic limb threatening ischaemia in Asian patients from Singapore – 12 months results of the PRISTINE registry. *Cardiovasc Intervent Radiol* (2024) 47:863–74. doi:10.1007/s00270-024-03756-3
115. M.A. Med Alliance S.A. SELUTION SLR™ 014 BTK. A prospective randomized multicenter single blinded study to assess the safety and effectiveness of the SELUTION

SLR™ 014 drug eluting balloon in the treatment of below-the-knee (BTK) atherosclerotic disease in patients with chronic limb threatening ischemia (CLTI). Clinical trial registration NCT05055297, clinicaltrials.gov (2025). Available online at: <https://clinicaltrials.gov/study/NCT05055297> (Accessed October 29, 2025).

116. Concept Medical Inc. *FUTURE BTK: randomized controlled trial of first Sirolimus Coated balloon versus Standard balloon angioplasty in the Treatment of below the knee arterial disease*. Clinical trial registration NCT04511247, clinicaltrials.gov (2025). Available online at: <https://clinicaltrials.gov/study/NCT04511247> (Accessed October 29, 2025).

117. Teichgräber U. *Prospective multi-center randomized controlled trial to evaluate the safety and efficacy of Sirolimus drug coated balloon versus non-coated standard angioplasty for the treatment of infrapopliteal occlusions in patients with Peripheral arterial Disease*. Clinical trial registration NCT04772300, clinicaltrials.gov (2024). Available online at: <https://clinicaltrials.gov/study/NCT04772300> (Accessed October 29, 2025).

118. Concept Medical Inc. *MAGICAL BTK: randomized controlled trial of MAGICTouch - sirolimus coated BALloon versus standard balloon angioplasty in the treatment of below the knee arterial disease*. Clinical trial registration NCT06182397, clinicaltrials.gov (2025). Available online at: <https://clinicaltrials.gov/study/NCT06182397> (Accessed October 29, 2025).

119. Atigh MK, Turner E, Christians U, Yazdani SK. The use of an occlusion perfusion catheter to deliver paclitaxel to the arterial wall. *Cardiovasc Ther* (2017) 35:e12269. doi:10.1111/1755-5922.12269

120. Zeller T, Zhang Z, Parise H, Mascho C, Holden A, Schmidt A, et al. *Early tibial vessel recoil following treatment with the bare temporary spur stent system: results from the DEEPER OUS vessel recoil substudy*. *J Endovasc Ther* (2024).

121. A novel temporary stent for treatment of infrapopliteal arteries in conjunction with drug-coated balloon angioplasty: the DEEPER pilot study | journal of CLI (2025) Available online at: <https://www.clijournal.com/article/novel-temporary-stent-treatment-infrapopliteal-arteries-conjunction-drug-coated-balloon> (Accessed 15 July 2025).

122. Schweiger L, Gütl K, Rief P, Reiter C, Janisch M, Weinberg I, et al. Retrievable scaffold therapy combined with sirolimus-coated balloon angioplasty for infrapopliteal artery disease: final results from the DEEPER LIMUS trial. *Cardiovasc Intervent Radiol* (2025) 48:297–303. doi:10.1007/s00270-025-03987-y

123. Health CD. *Esprit BTK everolimus eluting resorbable scaffold system – P230036*. FDA (2024). Available online at: <https://www.fda.gov/medical-devices/recently-approved-devices/esprit-btk-everolimus-eluting-resorbable-scaffold-system-p230036> (Accessed October 29, 2025).

124. Varcoe RL, DeRubertis BG, Kolluri R, Krishnan P, Metzger DC, Bonaca MP, et al. Drug-eluting resorbable scaffold versus angioplasty for infrapopliteal artery disease. *New Engl J Med* (2024) 390:9–19. doi:10.1056/nejmoa2305637

125. DeRubertis BG, Varcoe RL, Krishnan P, Bonaca MP, O'Connor DJ, Pin R, et al. Drug-eluting resorbable scaffold versus balloon angioplasty for below-the-knee peripheral artery disease: 2-year results from the LIFE-BTK trial. *Circulation* (2025) 152:1076–86. doi:10.1161/circulationaha.125.075080

126. Haude M, Wlodarczak A, Schaaf Rvan der, Torzewski J, Ferdinande B, Escaned J, et al. A new resorbable magnesium scaffold for *de novo* coronary lesions (DREAMS 3): one-year results of the BIOMAG-I first-in-human study (2025). doi:10.4244/EIJ-D-23-00326

127. Biotronik AG. *BIOTRONIK-Safety and Clinical Performance of the Drug Eluting Resorbable Coronary MAGnesium Scaffold System (DREAMS 3G) in the Treatment of Subjects With de novo Lesions in Native Coronary Arteries: BIOMAG-II: a Randomized Controlled Trial*. *Clin Trial Registration NCT05540223*, clinicaltrials.gov (2024). Available online at: <https://clinicaltrials.gov/study/NCT05540223> (Accessed October 24, 2025).

128. Schwartz LB, Orr G, Santos JD, Haig C. *Multi-element bioresorbable intravascular stent*. US11234844B2 (2022). Available online at: <https://patents.google.com/patent/US11234844B2/en?assignee=efemoral+medical&oq=efemoral+medical> (Accessed October 25, 2025).

129. Saito S, Nef HM, Webster M, Verheye S. DynamX sirolimus-eluting Bioadaptor versus the zotarolimus-eluting Resolute Onyx stent in patients with *de novo* coronary artery lesions: design and rationale of the multi-center, international, randomized BIOADAPTOR-RCT. *Cardiovasc Revascularization Med* (2023) 55:76–82. doi:10.1016/j.carrev.2023.05.010

130. Elixir Medical Corporation. *DynamX bioadaptor global post-market registry: clinical trial of the elixir medical DynamX coronary bioadaptor system (Bio-RESTORE)*. Clinical trial registration NCT06074549, clinicaltrials.gov (2025). Available online at: <https://clinicaltrials.gov/study/NCT06074549> (Accessed October 25, 2025).